

ГЕНЕТИКА GENETICS



DOI: 10.18413/2658-6533-2020-6-2-0-1

A novel heterozygous variant in exon 32 of the *CHD7* gene (c.6923C>T) in a Syrian family with Kallmann syndrome

Abdulsamad Wafa¹ , Faten Moassass¹ , Suher Almedani¹ ,
Thomas Liehr² , Kathleen Wilhelm² , Manar As'sad¹ ,
Sarah Knippenberg³ , Ralf Glaubitz³ , Rami A. Jarjour¹ ,
Walid Al Achkar¹ 

¹ Atomic Energy Commission of Syria,
17 Nissan St., Damascus, 6091, Syria

² Friedrich Schiller University Jena,
1 Fuerstengraben St., Jena, 07737, Germany

³ Amedes Genetics,
50 Georgstraße St., Hannover, 30159, Germany

Corresponding author: Walid Al Achkar (ascientific@aec.org.sy)

Abstract

Background: Kallmann syndrome (KS) and CHARGE syndrome (CS) are rare heritable disorders in which anosmia and hypogonadotropic hypogonadism co-occur. KS is genetically heterogeneous with at least eight genes being involved in its pathogenesis, whereas CS is caused by autosomal dominant mutations exclusively in *CHD7* gene. The majority of CS-cases are sporadic and only few familial cases have been reported. In these families, mosaicism in one parent, as well as parent-to-child transmission of a *CHD7* mutation, were described. **The aim of the study:** To report a paternal transmission of a variant in exon 32 of the *CHD7* gene (c.6923C>T) in a familial case originally suggested to be affected by KS. **Materials and methods:** Five genes associated with KS were analyzed using Sanger sequencing and MLPA in a 17-year-old male. **Results:** The heterozygous variant leading to a change of amino-acid serine at position 2,308 to leucine was found in father his three children. **Conclusion:** Overall this report confirms the existence of KS without CS symptoms, caused by a mutation in a gene reported pathogenic only in CS.

Keywords: heterogeneity; mutation; CHARGE syndrome; hypogonadotropic hypogonadism; anosmia

Acknowledgements: we thank Prof. I. Othman, the Director General of Atomic Energy Commission of SYRIA (AECS) and Dr. N. Mirali, Head of Molecular Biology and Biotechnology Department for their support. This work was supported by the AECS.

For citation: Wafa A, Moassass F, Almedani S, et al. A novel heterozygous variant in exon 32 of the *CHD7* gene (c.6923C>T) in a Syrian family with Kallmann syndrome. *Research Results in Biomedicine*. 2020;6(2):154-159. DOI: 10.18413/2658-6533-2020-6-2-0-1

Introduction. Idiopathic hypogonadotropic hypogonadism (IHH; OMIM 146110), one of the most commonly inherited forms of diminished functional activity of the gonads, results from deficient hypothalamic of gonadotropin releasing hormone (GnRH) release or action [1]. IHH patients present with absent or impaired sexual development due to sex-steroid-hormone deficiency, low serum levels of the pituitary gonadotropins follicle-stimulating hormone (FSH) and luteinizing hormone (LH), and infertility [1]. Kallmann syndrome (KS; OMIM 147950) is a combination of congenital hypogonadotropic hypogonadism (HH; OMIM 146110) and decreased/absent sense of smell [2]. Anosmia, or the inability to smell, is the result of olfactory bulb defects [3], whereas HH presents as absent or impaired pubertal maturation and is caused by GnRH deficiency [4]. KS accounts for about 50% to 60% of IHH [5] while forms of IHH present with normal olfaction (i.e. norm-osmic idiopathic HH: nIHH). KS is clinically and genetically very heterogeneous; phenotypic features may include additionally cleft lip/palate, hearing impairment, dental agenesis, limb anomalies, renal agenesis, and mirror movements [6]. The majority of KS cases (~60%) are sporadic; i.e. only one person is affected in the family. In familial KS, autosomal recessive, autosomal dominant, and X-chromosomal recessive inheritance have been described [7]. Oligogenic mode of inheritance has also been suggested [8, 9]. At present, mutations in eight genes explain approximately 25–35% of KS cases. Heterozygous loss-of-function mutations in the *CHD7* gene were identified in patients with nIHH, KS, and CHARGE syndrome (CS) [10, 11]. CS is a highly variable disorder in which congenital anomalies, multisensory impairment, and variable mental retardation can occur (OMIM 214800). CHARGE is an acronym for ocular coloboma, heart defects, choanal

atresia, retardation of growth and/or development, genital hypoplasia and ear anomalies combined with deafness [12]. HH and anosmia are present in the majority of patients with CS [13]. Recently, it was proven that HH and anosmia may co-occur in CS, too [14], which means that KS can be considered as part of the phenotypic spectrum of CS. *CHD7* mutations are found in more than 90% of patients with typical CS [15].

Here we report a family with one patient with symptoms resembling KS however lacking CS symptoms, and with a novel heterozygous variant in exon 32 of the *CHD7* gene (c.6923C>T) resulting in an amino acid exchange (p.Ser2308Leu).

Material and Methods

Clinical information

A 17-year-old male was the fifth child born to consanguineous Syrian healthy parents with a remarkable family history (delayed puberty in the children). At birth of the index patient his mother and father were 34 and 36 years old, respectively. The mother reported no history of infection during this pregnancy except for a slight hemorrhage at its beginning. After otherwise uncomplicated pregnancy and delivery, the index presented with micropenis and small testes (Tanner scale I, data not shown). At 17 years medical analyses showed an FSH (follicle stimulating hormone) level of 0.15 (normal value: 1.5 – 12.4 mIU/ml), a LH (luteinizing hormone) level of 0.10 (normal value: 1.4 – 8.6 U/ml) and a testosterone total level of 1.9 (normal value: 2-8 nmol/l). The patient had a normal male 46,XY karyotype and azoospermia without AZF-chromosome microdeletions (results are not shown). Subsequent normal height (170 cm) and weight (68 kg), no coloboma, no choanal atresia, and no cardiovascular malformations. The patient suffered from anxiety, was nervous and slightly autistic. The patient's older brother and sister (the second and third child of the family) also expired delayed puberty but disappeared spontaneously

in the boy and in the girl after hormonal treatment for 1.5-2 years. The remainder family members were clinically healthy. The study was approved by the ethical committee of the Atomic Energy Commission, Damascus, Syria. Informed consents were obtained from the father and other family members.

Genetic analysis

Genomic DNA was extracted from peripheral blood using QIAamp DNA Blood Mini Kit (Qiagen GMBH, Hilden, Germany). Sequencing of the following genes was performed: Kallmann 1 *KALI* (*ANOS*; NM_000216), Kallmann 2 *FGFR1* (NM_23110.2), Kallmann 3 *PROKR2* (NM_144773), Kallmann 4 *PROK2* (NM_001126128), Kallmann 5 *CHD7*

(NM_017780). Analysis was performed step-wise by Sanger-sequencing and analyses of deletions and duplications by Multiplex Ligation-dependent Probe Amplification (MLPA, MRC-Holland).

Results. No mutations, deletions or duplications were found in *KALI* (*ANOS*; NM_000216), *FGFR1* (NM_23110.2), *PROKR2* (NM_144773) or *PROK2* (NM_001126128). However, in *CHD7* (NM_017780) a novel heterozygous variant in exon 32 of the *CHD7* gene (c.6923C>T) resulting in an amino acid exchange (p.Ser2308Leu) was identified (Fig.) in patient, as well as in older 22 year old sister, 24 year old brother and the father (Tab.).

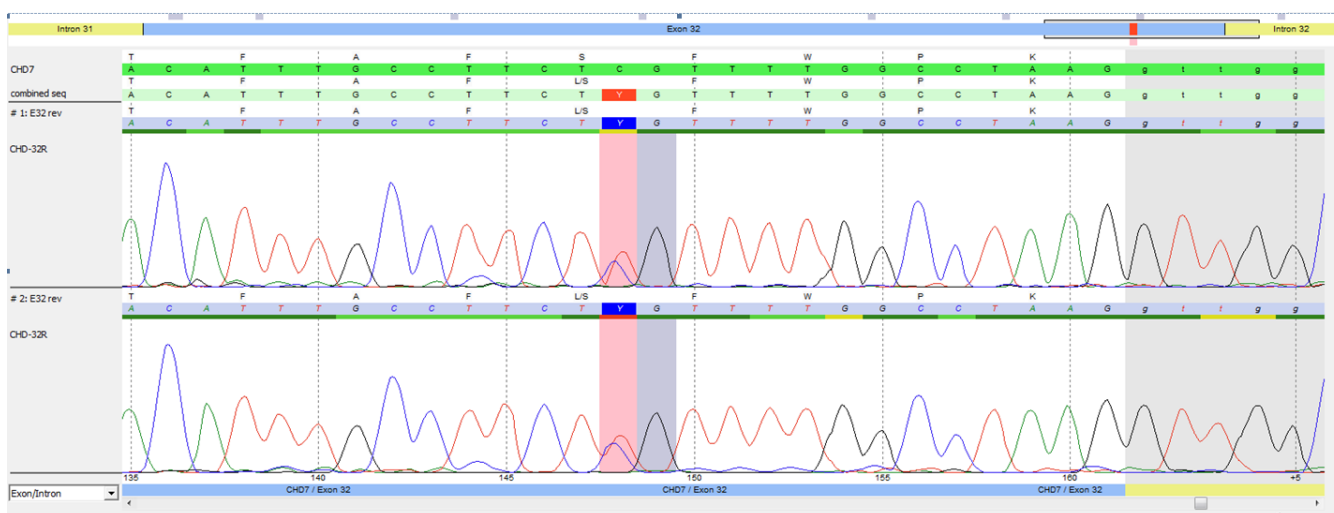


Fig. Results of analyses of the *CHD7*-gene, exon 32.

Analysis of the *CHD7*-gene sequence was performed by JSI Seq Pilot software. Heterozygous sequence-variation was detected at position c.6923 (c.6923C>T, p.Ser2308Leu)

Table

Status of the variant c.6923C>T in the family

No.	Family members	age	gender	Variant c.6923C>T
1	father	52	M	heterozygote
2	mother	50	F	absent
3	child 1	27	M	absent
4	child 2	24	M	heterozygote
5	child 3	22	F	heterozygote
6	child 4	20	F	absent
7	child 5	17	M	heterozygote

Discussion. KS is a unique form of IHH disease spectrum characterized by develop-

mental disorders with olfactory abnormalities being caused by congenital defects in GnRH

secretion of varying degrees. The pulsatile secretion of GnRH is essential for the hypothalamic-pituitary-gonadal axis function [16]. Also, KS is a unique disease model to study the migration of GnRH neurons and the development of human puberty. Some genes are necessary for the correct differentiation, migration, upstream signal regulation, and function of GnRH neurons in the embryonic period, which can lead to IHH [16]. Some genes that correctly differentiate embryonic GnRH neurons may be correlated with IHH, such as *KAL-1*, *FGFR1*, *FGF8*, *PROKR2*, *PROK2*, *CHD7*, *NELF*, *WDR11*, *HS6ST1*, *KISS1R*, *KISS1*, *TAC3*, *TACR3*, *LEPR*, *LEP*, *PCSK1*, *GNRHR*, *GNRH1*, *SEMA3A*, and *NDN7* [16]. Mutations in these genes lead to certain degrees of clinical manifestations. Moreover, KS can be caused by genes such as *KAL-1*, *FGFR1*, *PROKR2*, *PROK2*, *CHD7*, and *FGF8* [16]. Among them, the *CHD7* mutation was only found in KS patients with the CS phenotype, suggesting that if a patient was diagnosed with hypogonadism and anosmia, attention should be paid to the investigation of the presence of clinical characteristics of CS [16]. Here we report a paternally inherited new *CHD7* mutation in a KS patient.

The *CHD7* gene is located on chromosome 8q12.1, which encodes DNA-binding protein 7 of helicase in the chromatin region. This protein family has a unique functional domain binding site, including 2 N-terminal chromatin domains, 1 SWI2/SNF2-like ATP enzyme/solution helix domain, and 1 DNA binding domain. *CHD7* protein complex is expressed in the olfactory epithelium, hypothalamus, as well as the pituitary gland, suggesting that this protein may play an important role in the development of the olfactory bulb and GnRH neuron migration. The genetic pattern of *CHD7* gene has not yet been fully understood, and may follow autosomal dominant inheritance, with its mutations accounting for 6% of all IHH patients [17].

Jongmans et al. [10] identified 3/38 KS patients harboring de novo *CHD7* mutations (2 stop-mutations and 1 missense) whereas the nIHH patients were negative for *CHD7* variants. However, all 3 KS patients with

CHD7 variants, upon additional phenotypic review, universally exhibited major CS features. In contrast, Kim et al. [11] identified 7/56 IHH patients (3KS, 4 nIHH) harboring *CHD7* mutations (2 intronic mutations leading to exon skipping and 5 missense mutations), all of whom lacked major CS phenotypes, thus implicating *CHD7* allelic variants as a cause of both KS and nIHH forms of IHH without CS features. In view of these conflicting data as to whether *CHD7* mutations are capable of causing KS or nIHH without full CS, Bergman et al. [18] examined 36 KS patients in whom they identified 3 with *CHD7* mutations (2 nonsense, and 1 de novo missense). However, all three subjects displayed additional CS features, leading to their conclusion that *CHD7* mutations do not cause isolated IHH. Laitinen et al. [19] revealed no *CHD7* mutations in 30 Finnish KS patients. Jie et al. [16] found a family (two sons inherited a mutation from his mother, but the mother and his younger son did not show clinical features of KS) suffering from KS and some clinical features of CS. Pedigree verification can be achieved by *CHD7* gene mutation c.6571G>A. Hyung-Goo Kim et al. [11] examined 7/111 IHH/KS patients in whom they identified 7 *CHD7* mutations (two splice and five missense), three unrelated probands with KS and four unrelated probands with IHH with *CHD7* mutations, demonstrating that *CHD7* is involved in either IHH or KS. However, Hyung-Goo Kim et al. [11] suggested new evidence for a role of *CHD7* in the pathophysiology of both normosmic IHH and KS patients without a CS phenotype.

The molecular basis for 70%–75% of IHH/KS patients remains unknown [11]. To date, only *FGFR1* mutations have been reported to cause either nIHH families or KS families. Although a homozygous *PROKR2* deletion was seen in a single family comprising both normosmic and anosmic patients, this represents variable expressivity within the same family [11]. Interestingly, Hyung-Goo Kim et al. [11] found a one IHH and one KS patient, who both lack the CS phenotype, possess the same mutations (Ser834Phe and IVS65G/C) reported previously in patients with CS, further demonstrating the allelic re-

lationship of both syndromes. The KS patient with the IVS65G/C mutation does not fulfill Blake's criteria for CS, although she does have hearing impairment and cleft lip and palate. This also indicates that the effects of modifying genes may determine whether the patient has the more severe CS phenotype rather than the milder IHH/KS phenotype [11].

Overall, the family report confirms the existence of autosomal dominant inheritance of KS with lack of CS symptoms and a mutation in the yet only reported pathogenic gene involved in CS. This heterozygous variant in exon 32 of the *CHD7* gene (c.6923C>T), leading to a change of amino acid serine at position 2,308 to leucine devoid of any known functional domain, and therefore, it is unlikely to be a dominant negative form of the protein. Therefore, we hypothesize that this mutation represents a null allele, causing disease due to haploinsufficiency. This is in agreement with previous findings suggesting that a full genetic dosage is required for complete function of *CHD7* [18]. This novel variant in the present case was inherited from his father. Also, this variant was found in two subsequent children of his family. Moreover, this observation would draw the attention of the clinicians on the germline and familial responsible for the variable intrafamilial expression, suggesting a careful genetic counseling.

Financial support

No financial support has been provided for this work.

Conflict of interests

The authors have no conflict of interest to declare.

References

1. Bhagavath B, Podolsky RH, Ozata M, et al. Clinical and molecular characterization of a large sample of patients with hypogonadotropic hypogonadism. *Fertility and Sterility*. 2006;85(3):706-713. DOI: <https://doi.org/10.1016/j.fertnstert.2005.08.044>
2. Kallmann FJ, Schoenfeld WA, Barrera SE. The genetic aspects of primary eunuchoidism. *Am J Ment Def*. 1944;48:203-236.
3. Yousem DM, Turner WJ, Li C, et al. Kallmann syndrome: MR evaluation of olfactory system. *American Journal of Neuroradiology*. 1993;14(4):839-843.
4. Mitchell AL, Dwyer A, Pitteloud N, et al. Genetic basis and variable phenotypic expression of Kallmann syndrome: towards a unifying theory. *Trends in Endocrinology and Metabolism*. 2011;22(7):249-258. DOI: <https://doi.org/10.1016/j.tem.2011.03.002>
5. Bianco SDS, Kaiser UB. The genetic and molecular basis of idiopathic hypogonadotropic hypogonadism. *Nature Reviews Endocrinology*. 2009;5:569-576. DOI: <https://doi.org/10.1038/nrendo.2009.177>
6. Jones JR, Kemmann E. Olfacto-genital dysplasia in the female. *Obstetrics and Gynecology Annual*. 1976;5:443-466.
7. Pitteloud N, Quinton R, Pearce S, et al. Digenic mutations account for variable phenotypes in idiopathic hypogonadotropic hypogonadism. *Journal of Clinical Investigation*. 2007;117:457-463. DOI: 10.1172/JCI29884
8. Sykiotis GP, Plummer L, Hughes VA, et al. Oligogenic basis of isolated gonadotropin-releasing hormone deficiency. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;107(34):15140-15144. DOI: <https://doi.org/10.1073/pnas.1009622107>
9. Vaaralahti K, Raivio T, Koivu R, et al. Genetic overlap between holoprosencephaly and Kallmann syndrome. *Mol Syndromol*. 2012;3(1):1-5. DOI: <https://doi.org/10.1159/000338706>
10. Jongmans MC, van Ravenswaaij-Arts CM, Pitteloud N, et al. *CHD7* mutations in patients initially diagnosed with Kallmann syndrome: the clinical overlap with CHARGE syndrome. *Clinical Genetics*. 2008;75:65-71. DOI: <https://doi.org/10.1111/j.1399-0004.2008.01107.x>
11. Kim HG, Kurth I, Lan F, et al. Mutations in *CHD7*, encoding a chromatin-remodeling protein, cause idiopathic hypogonadotropic hypogonadism and Kallmann syndrome. *American Journal of Human Genetics*. 2008;83(4):511-519. DOI: <https://doi.org/10.1016/j.ajhg.2008.09.005>
12. Pagon RA, Graham JM Jr, Zonana J, et al. Coloboma, congenital heart disease, and choanal atresia with multiple anomalies: CHARGE association. *Journal of Pediatrics*. 1981;99(2):223-227. DOI: [https://doi.org/10.1016/S0022-3476\(81\)80454-4](https://doi.org/10.1016/S0022-3476(81)80454-4)
13. Jongmans MCJ, Admiraal RJ, van der Donk KP, et al. CHARGE syndrome: the pheno-

typic spectrum of mutations in the CHD7 gene. *Journal of Medical Genetics*. 2006;43:306-314. DOI: <http://dx.doi.org/10.1136/jmg.2005.036061>

14. Bergman JEH, Bocca G, Hoefsloot LH, et al. Anosmia predicts hypogonadotropic hypogonadism in CHARGE syndrome. *Journal of Pediatrics*. 2011;158(3):474-479. DOI: <https://doi.org/10.1016/j.jpeds.2010.08.032>

15. Bergman JEH, Janssen N, Hoefsloot LH, et al. CHD7 mutations and CHARGE syndrome: the clinical implications of an expanding phenotype. *Journal of Medical Genetics*. 2011;48:334-342. DOI: <http://dx.doi.org/10.1136/jmg.2010.087106>

16. Wen J, Pan L, Xu X, et al. Clinical data and genetic mutation in Kallmann syndrome with CHARGE syndrome: Case report and pedigree analysis. *Medicine*. 2018;97(27):e11284. DOI: [10.1097/MD.00000000000011284](https://doi.org/10.1097/MD.00000000000011284)

17. Dodé C, Hardelin JP. Clinical genetics of Kallmann syndrome. *Annales d'Endocrinologie*. 2010;71(3):149-157. DOI: <https://doi.org/10.1016/j.ando.2010.02.005>

18. Bergman JEH, Janssen N, van der Sloot AM, et al. A novel classification system to predict the pathogenic effects of CHD7 missense variants in CHARGE syndrome. *Human Mutation*. 2012;8:1251-1260. DOI: <https://doi.org/10.1002/humu.22106>

19. Laitinen EM, Vaaralahti K, Tommiska J, et al. Incidence, phenotypic features and molecular genetics of Kallmann syndrome in Finland. *Orphanet Journal of Rare Diseases*. 2011;6:41. DOI: <https://doi.org/10.1186/1750-1172-6-41>

Received 20 April 2020

Revised 27 May 2020

Accepted 2 June 2020

Information about the authors

Abdulsamad Wafa, BS, Laboratory Manager of Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: atomic@aec.org.sy, ORCID: 0000-0002-6246-9790.

Faten Moassass, BS, MS, Researcher of Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: atomic@aec.org.sy, ORCID: 0000-0002-4495-1866.

Suher Almedani Researcher of the Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: atomic@aec.org.sy, ORCID: 0000-0001-5179-0602.

Thomas Liehr, PhD, PD, Head of Molecular Cytogenetics Laboratory, Institute of Human Genetics of the Jena University Hospital, Friedrich Schiller University Jena, E-mail: Thomas.Liehr@med.uni-jena.de, ORCID: 0000-0003-1672-3054.

Kathleen Wilhelm, MD, Medical Doctor of the Institute of Human Genetics of the Jena University Hospital, Friedrich Schiller University Jena, E-mail: Thomas.Liehr@med.uni-jena.de, ORCID: 0000-0003-4153-9814.

Manar As'sad, Researcher of Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: atomic@aec.org.sy, ORCID: 0000-0002-5664-8960.

Sarah Knippenberg, Dr. rer. nat., Scientific Associate, Molecular Genetic of the Amedes Genetics, E-mail: info@amedes-genetics.de, ORCID: 0000-0002-0626-6016.

Ralf Glaubitz, MD, Molecular Genetic, Medical Director of the Amedes Genetics, E-mail: info@amedes-genetics.de, ORCID: 0000-0002-3643-1035.

Rami A. Jarjour, PhD, MD, Head of Laboratory of Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: atomic@aec.org.sy, ORCID: 0000-0003-3688-8809.

Walid Al Achkar, PhD, Head of Laboratory of Human Genetics Division, Molecular Biology and Biotechnology Department of the Atomic Energy Commission of Syria, E-mail: ascientific@aec.org.sy, ORCID: 0000-0002-1235-6990.